

Microplankton on Distribution in the South China sea, Area II: Sarawak, Sabah and Brunei Darussalam Waters

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ABSTRACT

Collaborative cruises in the South China Sea around the coast of Sarawak, Brunei and Sabah were conducted in the premonsoon (October, 1996) and the postmonsoon (June, 1997) periods on board MV SEAFDEC. The microplankton from 79 sampling stations consisted of more than 200 taxa consisting predominantly of blue green algae (3 species), diatoms (> 90 species), dinoflagellates (> 70 species) and microzooplankton (> 20 groups). Among the microplankton collected, three species of blue green (*Trichodesmium erythraeum*, *T. thiebautii*, *Richelia* sp.) and numerous diatom species were dominant. The dominant diatom species comprised of *Chaetoceros diversum*, *C. peruvianum*, *C. laciniosus*, *Thalassionema frauenfeldii*, *Bacteriastrum comosum*, *Coscinodiscus* sp. and *Rhizosolenia alata*; while those of dinoflagellates consisted of *Ceratium fusus*, *C. arcuatum*, *C. teres*, *Protoperidinium* sp., *Protoceratium* sp., *Ceratocorys* sp. and *Alexandrium* sp. The genera *Chaetoceros*, *Rhizosolenia*, *Bacteriastrum* and *Ceratium* were found to contain a wide range of species. The total microplankton densities ranged from 0.74×10^6 to 7.94×10^6 individuals / m³ and from 0.16×10^6 to 1.25×10^6 individuals / m³ during the premonsoon and postmonsoon periods respectively. The presence of the dinoflagellate species of *Ceratium*, *Protoperidinium* and *Alexandrium* were detected in considerable amounts at coastal and intermediate middle waters of the South China Sea. Blooms of *Rhizosolenia alata* and *Trichodesmium* sp. occurred during the premonsoon period. The microzooplankton consisted of more than 20 species dominated by copepod nauplii (> 50% of total microzooplankton count), radiolarians, foraminiferas and protozoans; most of the zooplankton species were dominant in nearshore and intermediate middle waters of the South China Sea.

Introduction

Published works on studies of plankton and other related organisms of the Malaysian waters in the South China Sea are scanty. Qualitative studies of plankton in the Malaysian coastal waters, especially the Malacca Straits have been conducted by Sewell (1933), Wickstead (1961) and Pathansali (1968). Primary productivity in the same location had been carried out by Doty *et al.* (1963); however, a detailed study of the species community structure, distribution and abundance of plankton in such waters had been lacking. Studies by Shamsudin (1988) in the South China Sea around coasts of Johore, Terengganu and Kelantan found that majority of the phytoplankton found were diatoms which comprise of numerous species of *Bacteriastrum*, *Chaetoceros*, *Rhizosolenia* and *Pleurosigma*. The blue green, *Trichodesmium erythraeum* was found in abundance in tropical waters (Chua & Chong, 1973). Studied on plankton (Shamsudin 1987, Shamsudin & Baker 1987, Shamsudin *et al.* 1987, Chua & Chong 1973) had raised questions about the qualitative and quantitative seasonal availability of these organisms as sources of food for those organisms higher up in the food chain and the relative production of these organisms in various study sectors of the South China Sea.

Studies by Chua and Chong (1973) in the Malacca Straits showed that the distribution and abundance of pelagic species especially the small tuna (*Euthynus affinis*), chub makerel (*Rastrelliger* sp.) and anchovies (*Stolephorus* sp.) were related to the density of phytoplankton. Physico-chemical factors may influence the distribution and abundance of phytoplankton. These factors consisted of temperature, salinity, dissolved oxygen, turbidity, current and nutrient concentration (nitrate, phosphate, ammonium and silicate).

In the present study, the microplankton community structure has been analysed during the pre and postmonsoon periods (October 1996, June 1997) in the Sarawak, Sabah and Brunei waters of the South China Sea. The species community structure patterns, distribution, composition and species abundance at various study sectors of the South China Sea during the two seasons were estimated.

Methods

Study Area

The study area covers an area which extends from the eastern Sarawak waters (Lat. 2° 18.4' E; Long. 109° 35.4' N) to the northern tip of Sabah covering the Sabah waters (7° 9.6' E; 115° 51.9' N) of the South China Sea. The estimated study area is ca 8100 nautical square miles (ca 25100 sq. km) covering the economic exclusive zone (EEZ) of Malaysian sea of the South China Sea. The sea cruise track followed a zig-zag manner starting from the eastern coastal Sarawak waters and ended up at the northern end of Sabah waters covering a total of 79 sampling stations.

Sampling Method & Preparation

The research survey were carried out in October 1996 and June 1997 whereby seventy nine stations were identified during the survey. Vertical plankton net of mesh size 56 µm was hauled at a speed of 1 m/s from 40 m (twice the depth of the 1% surface illumination) to the surface. The net diameter is 45 cm and length 92 cm. Samples at various depths using Van Dorn water sampler (20 litres) were also taken to quantify the microplankton population which also include some of the microzooplankton. This is to compensate the error which might arise from plankton escaping the net plankton. During sampling the plankton also included some zooplankton that might be caught in the water sampler and net plankton at the same time. The net plankton was hauled vertically at very slow speed (to avoid plankton avoidance of the plankton net) at every stations. The samples were preserved in 10% formalin. The microplankton fractions of the samples were examined for species composition and abundance. A quantitative study of the microplankton was carried out using an inverted microscope (Vollenweider *et al.* 1974; Tippett 1970; Shamsudin 1987, 1993, 1994, 1995; Shamsudin & Shazili 1991; Shamsudin & Sleight 1993, 1995; Shamsudin *et al.* 1987, 1997).

The microplankton cells were routinely examined with a Nikon microscope using a x 10 eyepiece and a x 40 bright field objective. Difficult specimens were examined under a x 100 oil immersion objective. Where it was necessary for a detailed identification, samples were treated by boiling and washing in 10% HCl (Tippett, 1970) to clean diatom frustules in order to show up their ultra fine structure for identification purposes, employing the scanning electron microscope (SEM) technique. The samples which had been fixed and preserved in absolute alcohol, were then mounted on (SEM) stubs with double-sided cellotape. The stubs with adhering samples were then coated with an alloy (gold with palladium) before being observed under the scanning electron microscope (Barber & Haworth, 1981). Algal were identified with reference to Palmer & Keely (1900), Cleve (1901, 1904), Gran (1912), Pascher (1914, 1915 & 1925), Hustedt (1930), Sewell (1933), Handey (1933, 1964), Fritsch (1935), Cummins & Mulryan (1937), Cupp (1943), Cleve-Euler (1944), Crossby & Wood (1959), Winstead (1961), Banse (1964), Patrick & Reimer (1968), Shiota (1966), Newell & Newell

(1973), Taylor (1976), Taylor & Seliger (1979) and Barber & Haworth (1981).

An index of the composition of the plankton community in the aquatic habitat is given by calculating the diversity index (H) and evenness (J) of the community structure using the Shannon-Weiner (1949) index. The formula for calculating Shannon-Weiner (diversity) index (H) is :

$$H = -\sum P_i \log_2 P_i, \text{ Where } P_i = n_i/N$$

n_i = The number of individuals of the i th species

N = The total number of individuals

The diversity index can measure species richness (H) and species evenness (J)

$$J = H/\log_2 S - (ii), \text{ S is the number of species}$$

Statistical Analysis

Analysis of variance can be used to assess the relative importance of different sources of variation, e.g. between sites, between dates, etc., but it may be necessary to transform the data before analysis of variance tests are applied. One way analysis of variance can be employed when comparisons are made between a number of independent random samples, one sample from each population. All counts must be classified in the same manner, but the number of counts in the various samples can be different (Elliott, 1977).

Coefficients of similarity are simple measures of the extent to which two habitats have species (or individuals) in common (Southwood, 1978). Essentially, such coefficient can be of two types, as given below, and both types reflect the similarity in individuals between the habitats.

(i) Jaccard $C_j = j / (a + b - j)$

(ii) Sorensen $C_s = 2j / (a+b)$

where a , b are the total individuals sampled in habitat a and b respectively, and j is the sum of the lesser values for the species common to both habitats (Southwood, 1978). In habitats where one or few species have high dominance the coefficients under-estimate the contributions of the moderately common species which may be more stable indicators of the characteristic fauna of an area while the rare species have little impacts (Southwood, 1978). It is apparent that C_s is greater than C_j and the inequality reduces as j approaches the magnitude of $1/2 (a+b)$.

The microplankton can be classified into species assemblages or associations in cluster analysis on species sampled from the nearshore and offshore stations according to their preference on environmental conditions using the unweighted pair group average (UPGA) Pearson correlation index (Pielou, 1984; Ludwig & Reynolds, 1988).

Results and Discussion

The microplankton during the pre and post monsoon survey cruises consisted of more than 200 taxa consisting predominantly of blue green algae (3 species), diatoms (> 90 species) and dinoflagellates (> 70 species) (Tables 1 & 2, see Appendix). Three species of blue greens (*Trichodesmium erythraeum*, *T. thiebautii*, *Richelia* sp.) as well as several species of diatoms and dinoflagellates were dominant. The dominant diatom species comprised of *Chaetoceros diversum*, *C. peruvianum*, *C. laciniosus*, *Thalassionema frauenfeldii*, *Rhizosolenia alata*, *R. hebatata*, *R. styliformis*, *Bacteriastrum comosum*, *B. varians*, *B. hyalimum*, *Coscinodiscus* sp. and *Rhizosolenia alata*; while those of dinoflagellates consisted of *Ceratium fusus*, *C. teres*, *C. areuatum*, *Protoperidinium* sp., *Protoceratium* sp., *Ceratocorys* sp. and *Alexandrium* sp (Tables 2 & 3). The genera *Chaetoceros*, *Rhizosolenia*, *Coscinodiscus*, *Bacteriastrum* and *Ceratium* were found to contain a wide range of species. The total microplankton densities ranged from 0.74×10^6 to 7.94×10^6 individuals / m^3 and from 0.16×10^6 to 1.25×10^6 individuals / m^3 during the premonsoon and postmonsoon periods respectively (Fig. 1). There was an increase of about one order of magnitude in the total cell population during the premonsoon as compared to the post monsoon season. The diversity index H values ranged from 1.7 to 4.8 with usually high values in the coastal stations during both seasons (Fig. 2). The J evenness index values were usually directly proportional to the H values.

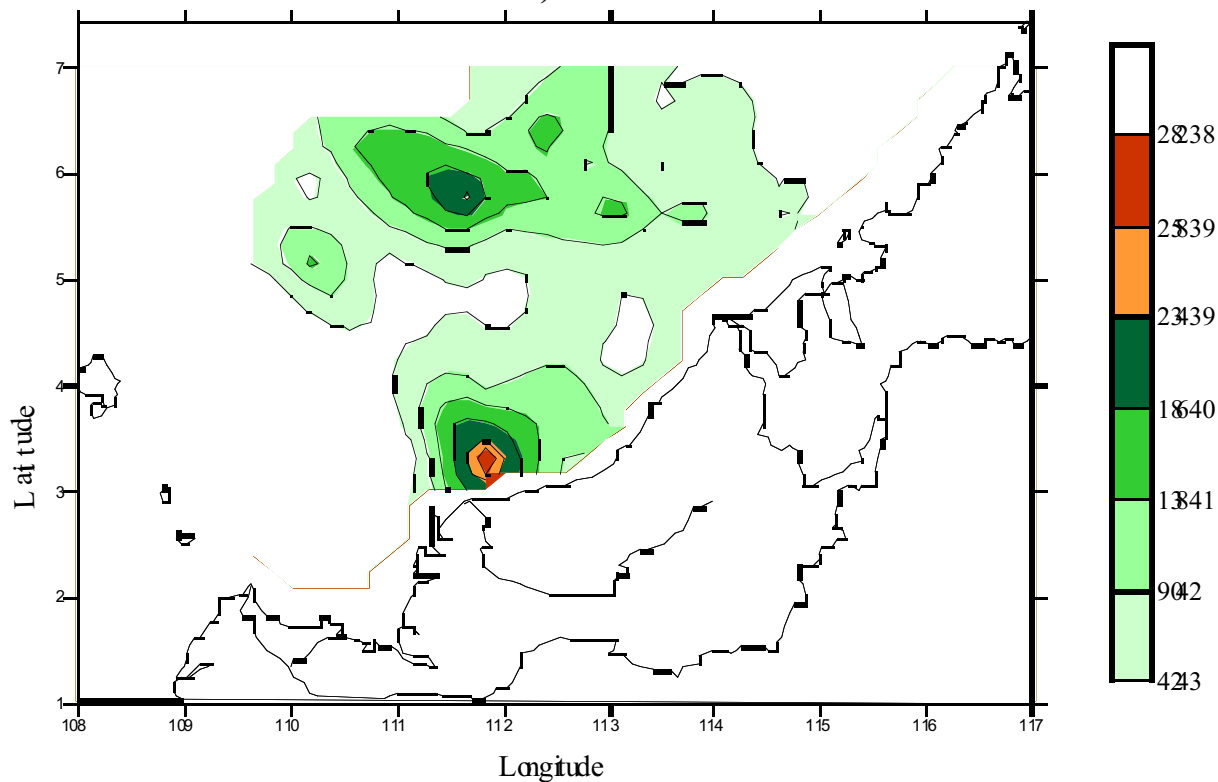
The maps from Figs. 3.1 a & b show the total cell population density during the pre and post monsoon period during the cruise survey with lesser population density in the latter. Two distinct patches of high cell population density were observed near the Rajang river mouth and offshore waters to the north of Sarawak during the premonsoon. A broad strip of water mass rich in microplankton was observed in the middle Sarawak waters of the South China Sea during the postmonsoon. It is noted that two patches of *Trichodesmium* bloom occurred near the Rajang river mouth and offshore waters to the north of Sarawak (Figs. 3.2 a & b). The dinoflagellate bloom was dense at the Rajang river mouth during the pre monsoon period while it was scattered during the postmonsoon (Figs. 3.3 a & b). An elongated narrow strip of *Thalassionema* bloom was observed along the coast of Sarawak during the premonsoon while two small patches of *Thalassionema* bloom was seen during the postmonsoon (Figs. 3.4 a & b). An isolated *Rhizosolenia* bloom was seen in the middle waters to the north of Brunei during the postmonsoon (Figs. 3.5 a & b). The species of *Trichodesmium* and *Rhizosolenia* were dominant species during the pre and post monsoon periods; numerous small patches of different species of *Chaetoceros*, *Thalassionema* and *Bacteriastrum* are also found (Figs. 3.6 a & b) during the study period.

Microplankton population at various sectors

Sampling stations can be grouped into at least 5 sectors with respect to their similarities in species composition using cluster analyses on 79 stations by mean of the unweighted pair group average (UPGA) Pearson index analyses (Fig. 4). The identified sectors comprised of a) Sarawak coastal waters (SCW), b) Sarawak middle water (SMW), c) Offshore Sarawak waters (OSW), d) Eastern Sabah waters (ESW) and e) Western Sarawak waters (WSW). The mean population densities at various stations of the 6 sectors (data from various stations from each sector were pooled together) were high during the premonsoon with values ranging from 0.74×10^6 to $7.94 \times 10^6 / m^3$ with Sarawak coastal waters sector having the highest values (Figs. 5 - 9). The trend in the mean densities at the 5 sectors during the post monsoon was similar to that of the premonsoon; however the values were much lower during the postmonsoon, ranging from 0.16×10^6 to $1.25 \times 10^6 / m^3$.

The major microplankton species at the Sarawak middle waters sector during premonsoon comprised of *Rhizosolenia calcar-avis*, *Chaetoceros lorenzianus*, *Coscinodiscus* sp., *Trichodesmium erythraceum* and *Ceratium fusus* with values ranging from 794 to $9.7 \times 10^5 / m^3$. The copepod nauplii had a moderate value of $6.31 \times 10^4 / m^3$ (Fig. 5). Microplankton species during the premonsoon were

Oct. 96
a) Total / l



June 97
b) Total / l

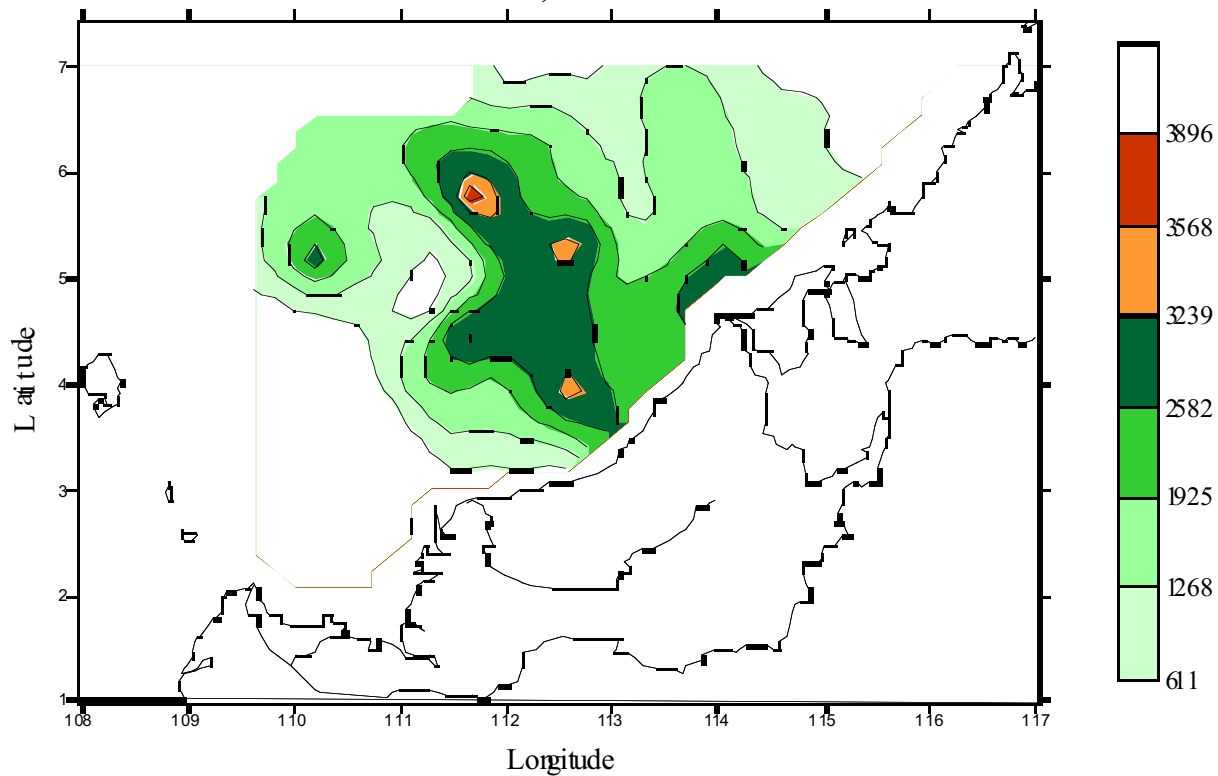


Fig. 3.1 Total cell density (L⁻¹) of microplankton cell population during the (a) pre and (b) post monsoon period of the cruise survey (October 1996 and June 1997).

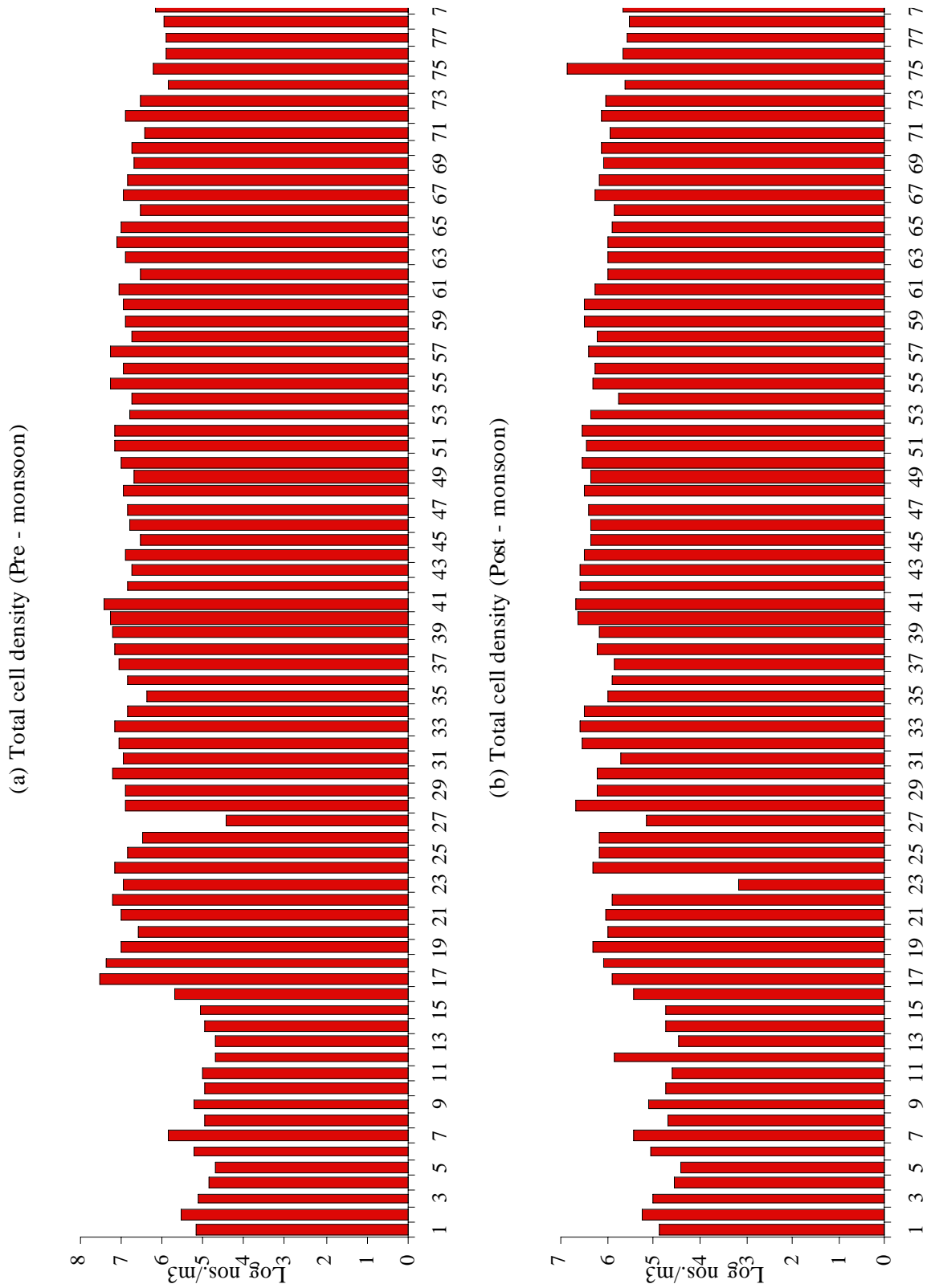


Fig. 1 Total cell densities (log nos./m³) at different stations during the pre and post - monsoon period (Oct. 1996 / June 1997 respectively) in Sabah - Sarawak waters of the South China Sea.

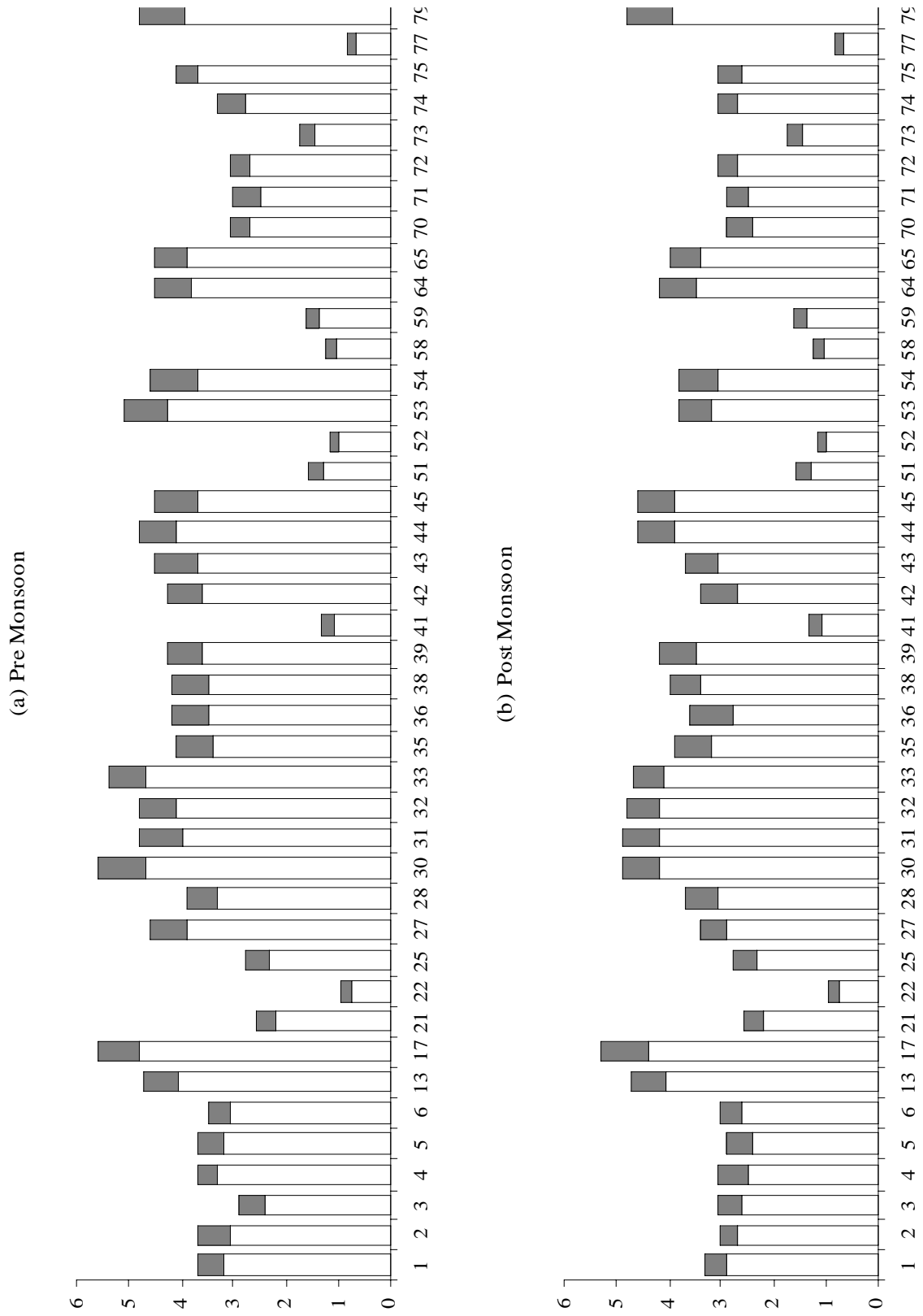
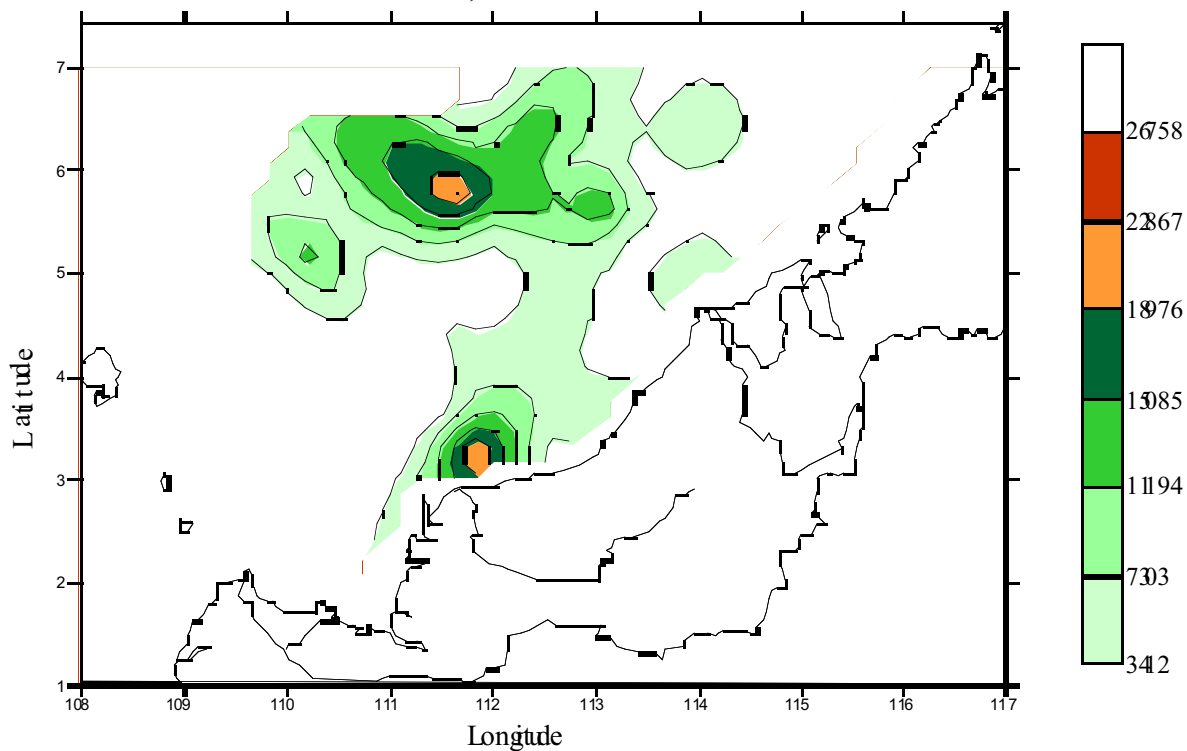


Fig. 2 The diversity (H) and evenness (J) indices of various station in the Sarawak - Sabah waters of the South China Sea during (a) pre monsoon and (b) post monsoon season.

Oct. 96
a) *Trichodesmium* / l



June 97
b) *Trichodesmium* / l

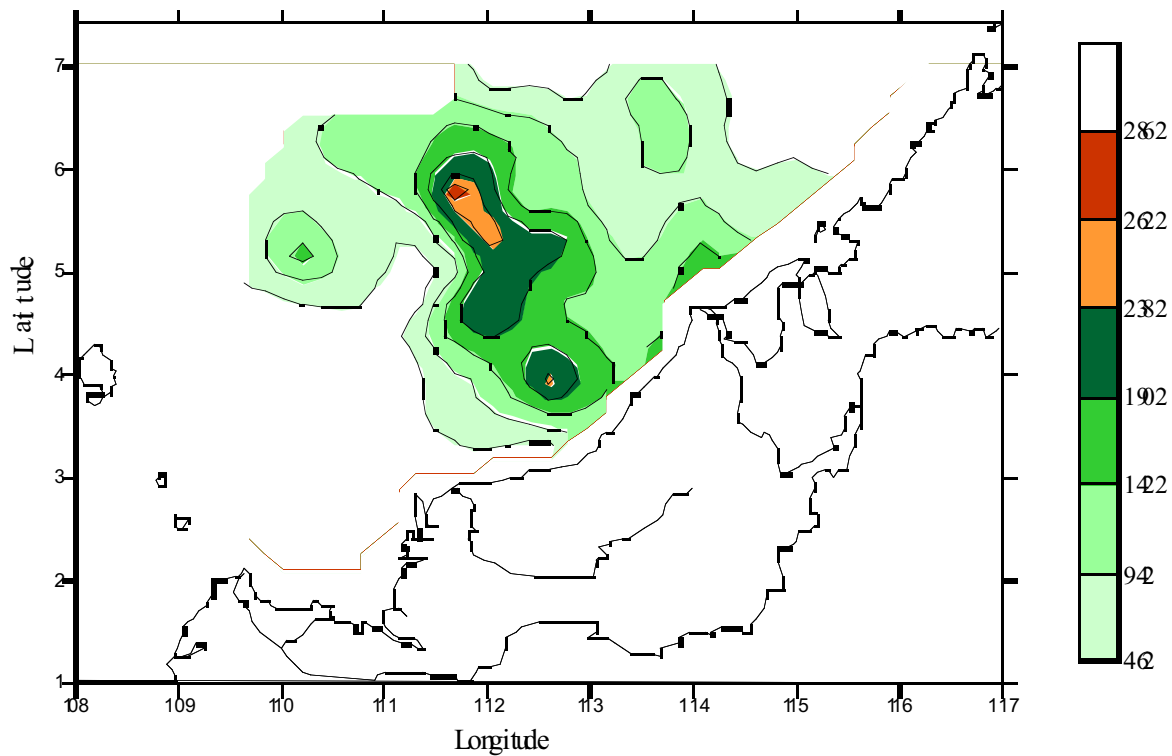


Fig. 3.2 *Trichodesmium* population density (L⁻¹) during the (a) pre and (b) post monsoon period of the cruise survey (October 1996 and June 1997).

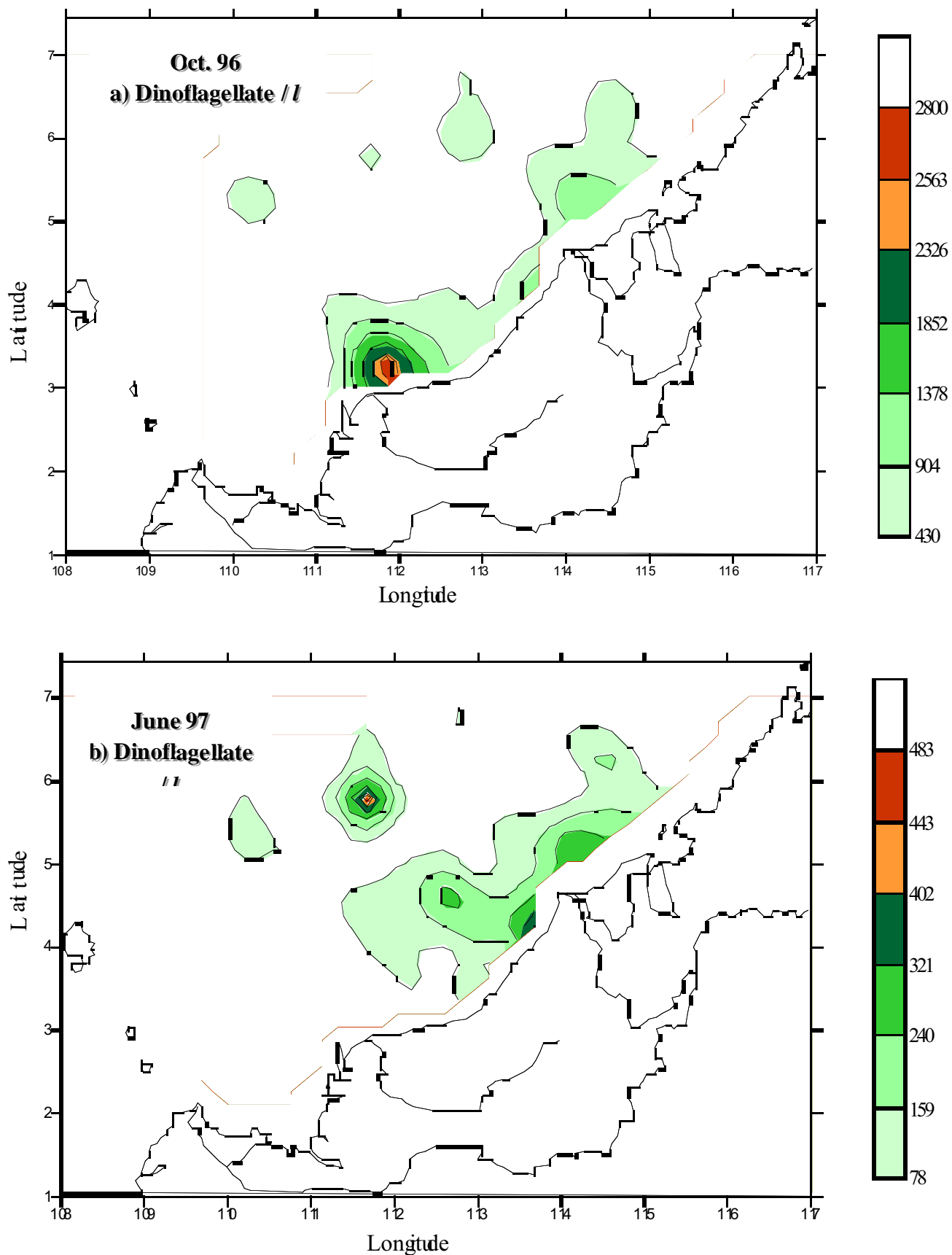


Fig. 3.3 Dinoflagellate population density (L^{-1}) during the (a) pre and (b) post monsoon period of the cruise survey (October 1996 and June 1997).

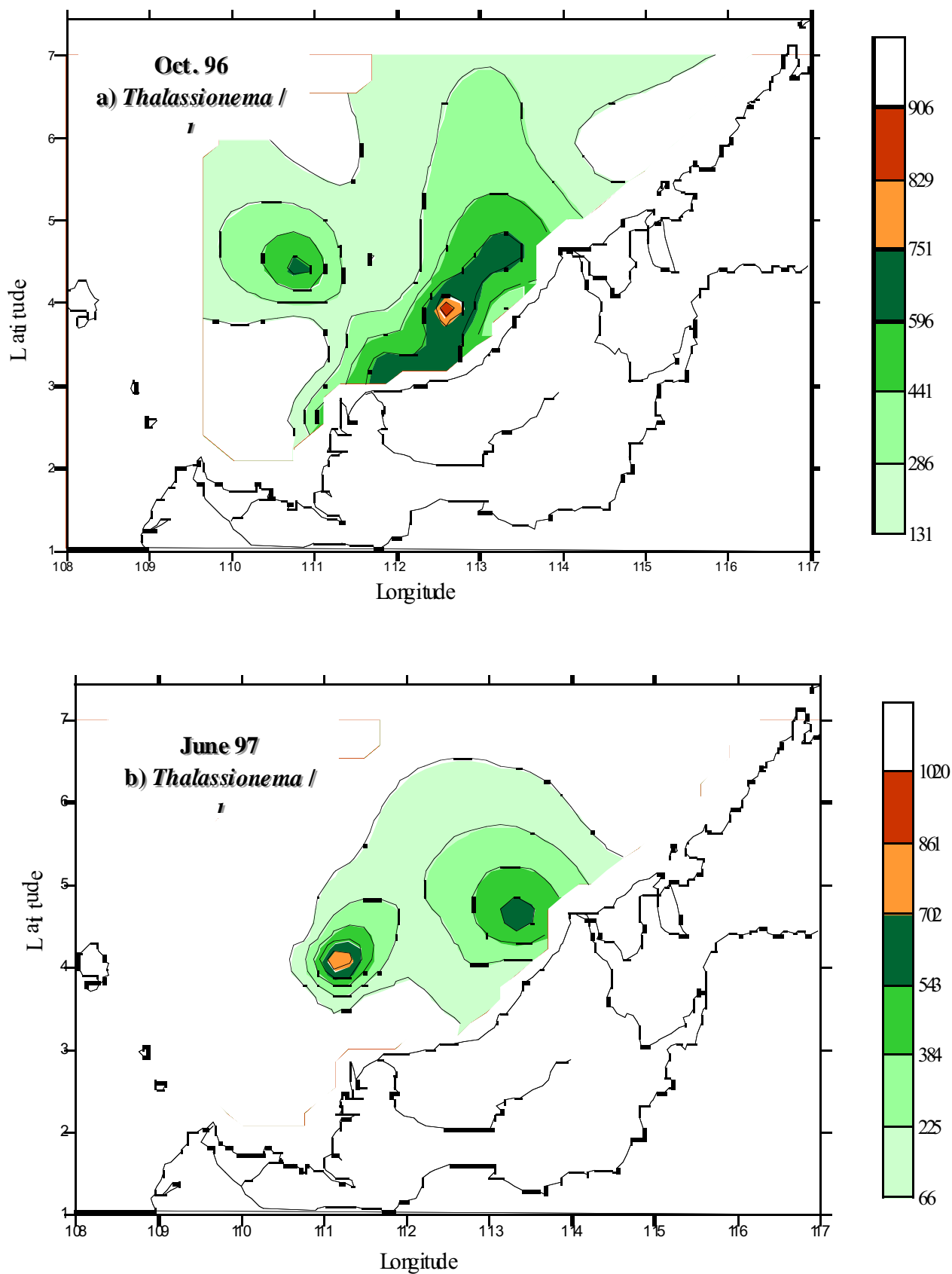


Fig. 3.4 *Thalassionema* population density (L^{-1}) during the (a) pre and (b) post monsoon period of the cruise survey (October 1996 and June 1997).

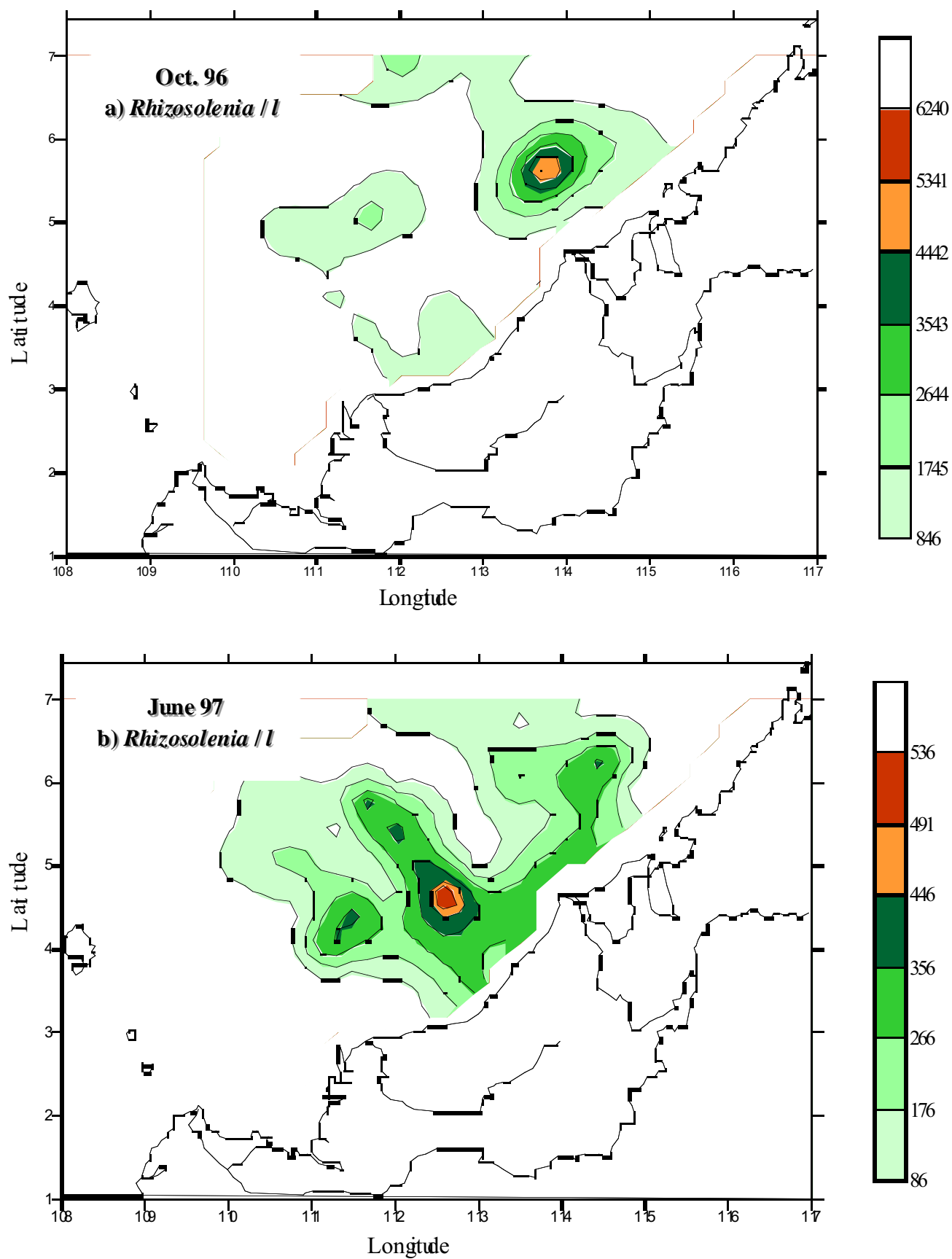


Fig. 3.5 *Rhizosolenia* population density (L^{-1}) during the (a) pre and (b) post monsoon period of the cruise survey (October 1996 and June 1997).

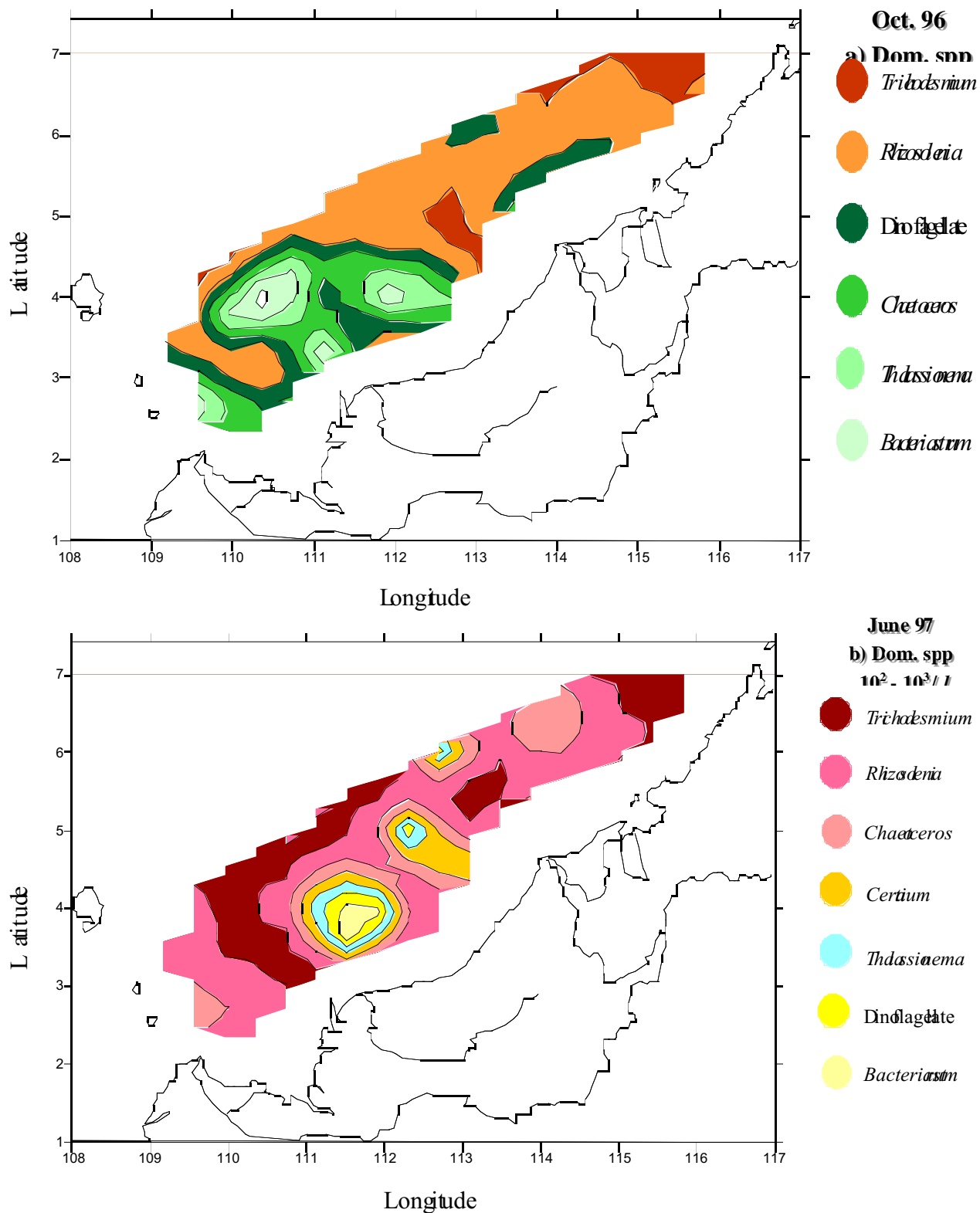


Fig. 3.6 Dominant microplankton species during the (a) pre and (b) post monsoon period of the cruise survey (October 1996 and June 1997).

present in higher concentrations than those during the postmonsoon.

The mean total population densities in the Sarawak middle waters were 5.01×10^6 and $7.9 \times 10^5/m^3$ during the pre and postmonsoon seasons respectively (Fig. 6). During the premonsoon the blue green, *Trichodesmium erythraeum* reached its peak bloom at concentration of $1.99 \times 10^6/m^3$ (> 30% of its total cell density); however, this species was again detected during the postmonsoon with a lower value ($6.3 \times 10^5/m^3$). *Rhizosolenium hebatata*, *Bacteriastrum* sp., *Chaetoceros lorenzianum*, *Thalassionema frauenfeldii* were dominant diatoms present during the premonsoon with values ranging from 1.9×10^3 to $2.51 \times 10^5/m^3$. *Coscinodiscus* sp. and *Thalassionema frauenfeldii* were present only during the premonsoon. Four species or *Rhizosolenia* (*R. alata*, *R. hebatata*, *R. styliformis* and *R. calcar - avis*) were also present with values ranging from 3.98×10^4 to $5.01 \times 10^5/m^3$. *Tintinnopsis* sp. and copepod nauplii were high during the premonsoon.

Trichodesmium bloom (comprising of *T. erythraeum* and *T. thiebautii*) occurred in offshore Sarawak waters with its peak density value of $2.24 \times 10^6/m^3$ (>20% of total cell density) (Fig. 7). The bloom occurred during both monsoons. *Rhizosolenia hebatata* was the only dominant diatom present during the premonsoon. Species of *Ornithocerus* and *Peridinium* were present with values ranging from 6.31×10^4 to $11.5 \times 10^4/m^3$. Fair amounts of *Richelia* sp., *Climacodium* sp., *Ceratium macroceros* and *C. fusus* also occurred during the two seasons. The mean total cell density of the post monsoon was 28% that of the premonsoon.

Chaetoceros macroceros and *Trichodesmium erythraeum* were dominant species in the eastern Sabah waters with mean densities of $1.58 \times 10^5 /m^3$ and $3.54 \times 10^5/m^3$ respectively; however both the species were present in lower concentrations during the postmonsoon (Fig. 8). *Protocentrum* sp., *Dinophysis* sp. and *Thalassionema nitzschoides* were present during the postmonsoon; however *Chaetoceros macroceros*, *Ceratium furca* and copepod nauplii were found during both seasons. *Protoperidinium* sp. was present in considerable concentration during the post monsoon. The mean total cell densities in this sector were 1.58×10^6 and $2.50 \times 10^5/m^3$ during the pre and postmonsoon seasons respectively.

The fifth sector was identified for those stations around the western nearshore Sarawak waters. During the premonsoon, the diatoms (*Rhizosolenia alata*, *Chaetoceros lorenzianum*, *Bacteriastrum delicatulum*) and the blue greens (*Trichodesmium erythraeum* and *T. thiebautii*) were dominant with values ranging from 2.51×10^3 to $1.38 \times 10^3/m^3$ (Fig. 9). Copepod nauplii were present during both seasons with values ranging from 150 to $1150/m^3$. The mean total cell densities were 1.99×10^5 and $1.58 \times 10^5/m^3$ during the pre and postmonsoon seasons respectively.

Microplankton assemblages and associations

The results from Fig. 10 illustrate that the microplankton species during the premonsoon comprised of at least seven species assemblages or associations in cluster analysis on 57 species sampled from the nearshore and offshore stations according to their preference on environmental conditions using the unweighted pair group average (UPGA) Pearsons index analyses. The species assemblages consisted of group A (*Chaetoceros coarctatum*, *Thalassionema fraeufeldii*, *Bacteriastrum delicatulum*); group B (*Trichodesmium erythraeum*, *Ceratium arcuatum*); group C (*Thalassionema fraeunfeldii*, *Chaetoceros lorenzianum*); group D (*Rhizosolenia hebatata*, *Chaetoceros laciniosus*, *C. decipiens*, *Ceratium longissimum*); group E (*Rhizosolenia styliformis*, *R. robusta*, *R. bergonii*, *R. alata*); group F (*Thalassionema thiebautii*, *Chaetoceros peruvianum*, *Ceratium teres*, *C. fusus*)(Table 3).

During the post monsoon period, cluster analysis of 51 species sampled from the 79 stations shows that there was a change in species assemblages or associations. At least 7 species associations according to their preference on environmental conditions (Fig. 11, Table 3). The species assemblages consisted of group A (*Ceratocorys* sp., *Bacteriastrum hyalinum*, *Nitzshia frigida*); group B (*Thalassionema fraeunfeldii*, *Nitzschia seriata*, *Rhizosolenia stolterforthii*, *Chaetoceros lorenzianum*);

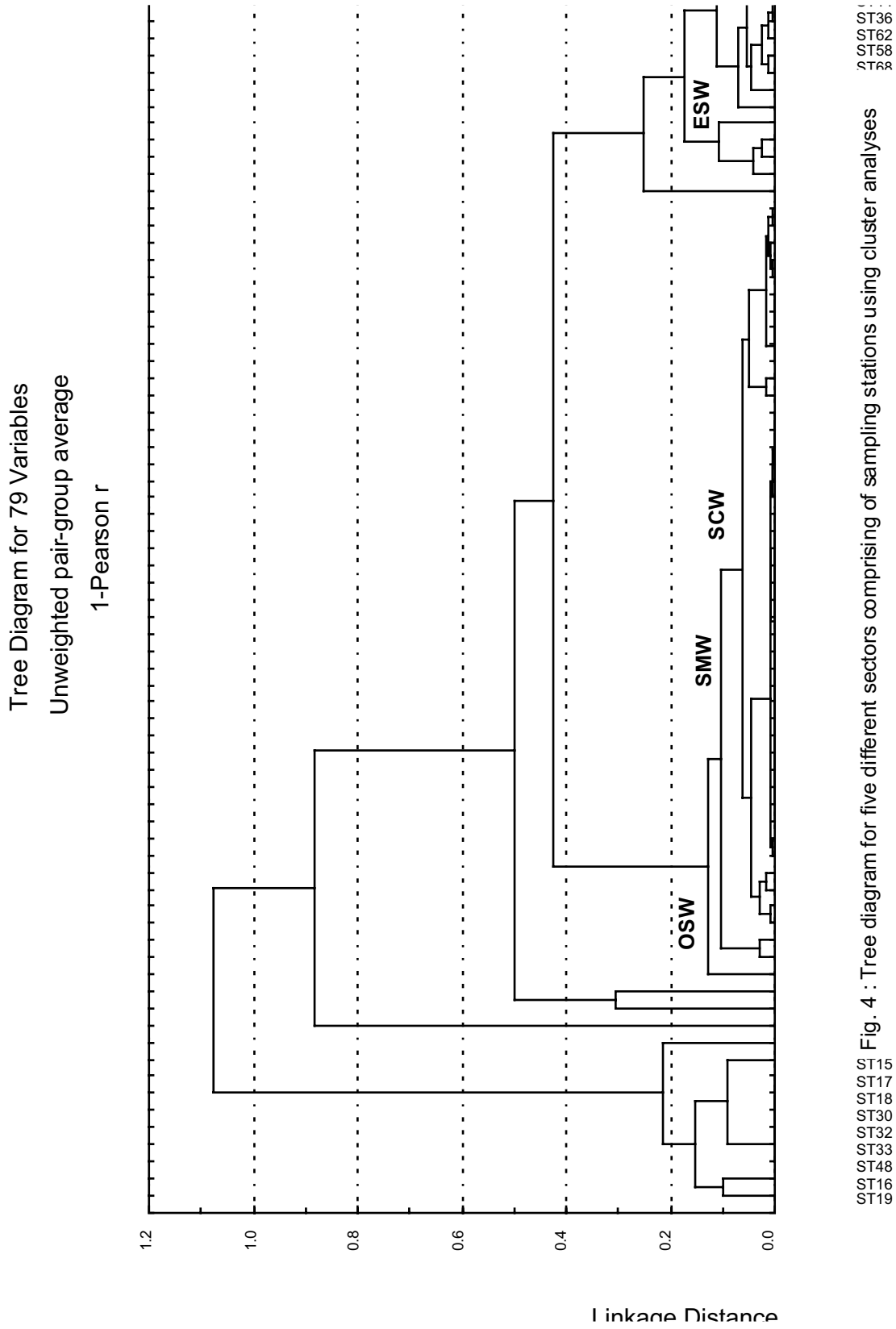


Fig 4

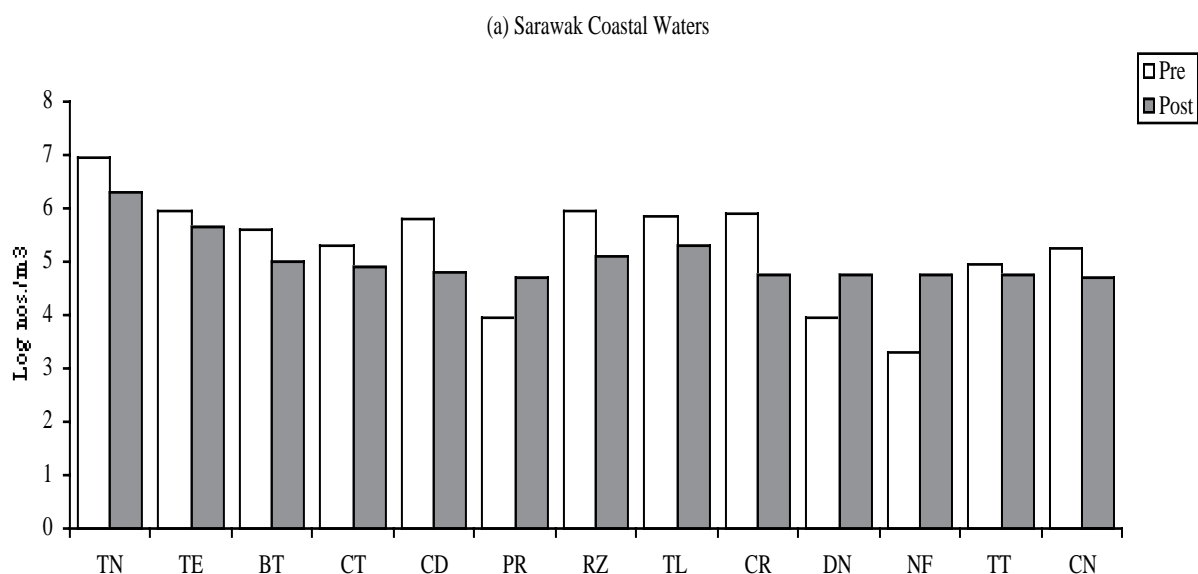


Fig. 5 Cell densities (log nos./m³) of various microplankton species at stations near Sarawak Coastal Waters during pre and post monsoon seasons. (TN-total cell, TE-*Trichodesmium erythraeum*, BT-*Bacteriastrum cosmosum*, CT-*Chaetoceros lorenzianum*, CD-*Coscinodiscus debilis*, PR-*Peridinium* sp., RZ-*Rhizosolenia calcar-avis*, TL-*Thalassionema frauenfeldii*, CR-*Ceratium fusus*, DN-*Dinophysis* sp., NF-*Nitzchia frigida*, TT-*Tintinnopsis* sp., CN-Copepod nauplii).

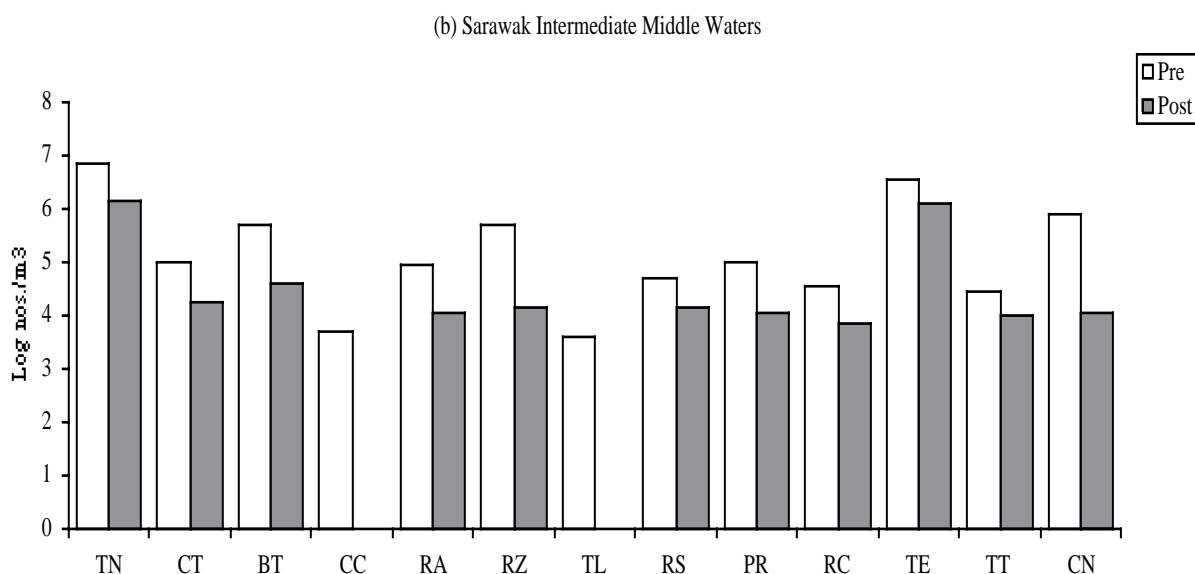


Fig. 6 Cell densities (log nos./m³) of various microplankton species at stations in Sarawak Intermediate Middle Waters of the South China Sea during pre and post monsoon seasons. (TN-total cell, CT-*Chaetoceros lorenzianum*, BT-*Bacteriastrum delicatulum*, CC-*Coscinodiscus* sp., RA-*Rhizosolenia alata*, RZ-*Rhizosolenia hebatata*, TL-*Thalassionema frauenfeldii*, RS-*Rhizosolenia styliformis*, PR-*Peridinium* sp., RC-*Rhizosolenia calcar-avis*, TE-*Trichodesmium erythraeum*, TT-*Tintinnopsis* sp., CN- Copepod nauplii).

(c) Offshore Sarawak Waters (OSW)

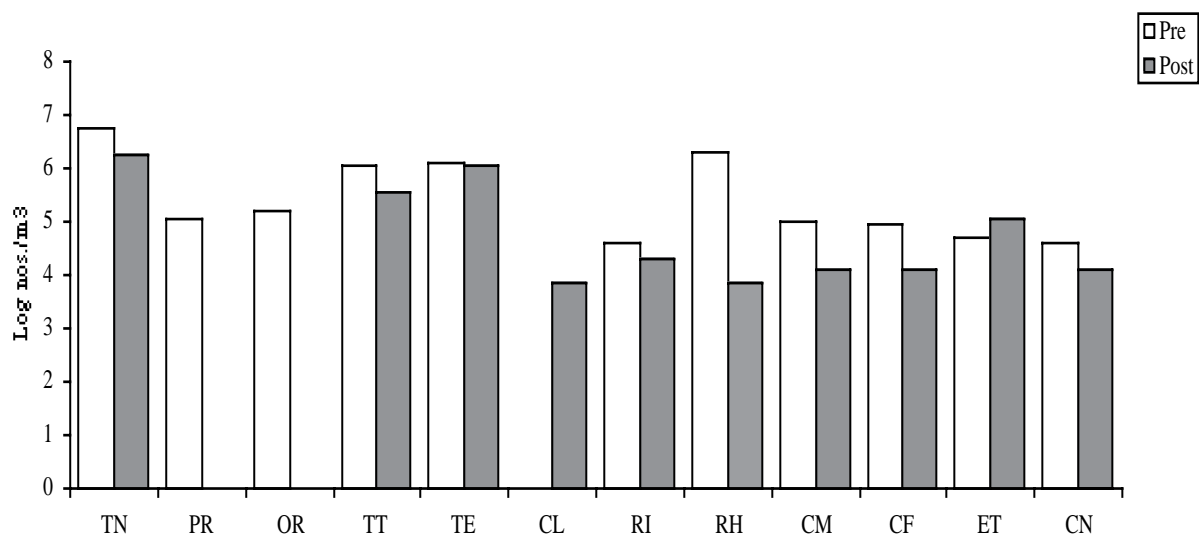


Fig. 7 Cell densities (log nos./m³) of various microplankton species at stations in Offshore Sarawak waters of the South China Sea during pre and post monsoon seasons. (TN. total cell, PR-*Peridinium* sp., OR-*Ornithocerus* sp., TT-*Trichodesmium thiebautii*, TE. *Trichodesmium erythraeum*, CL-*Climacodium* sp., RI-*Richelia* sp., RH-*Rhizosolenia hebatata*, CM-*Ceratium macroceros*, CF. *Ceratium fusus*, ET-*Eutintinus* sp., CN. Copepod nauplii)

(d) Eastern Sabah Waters (ESW)

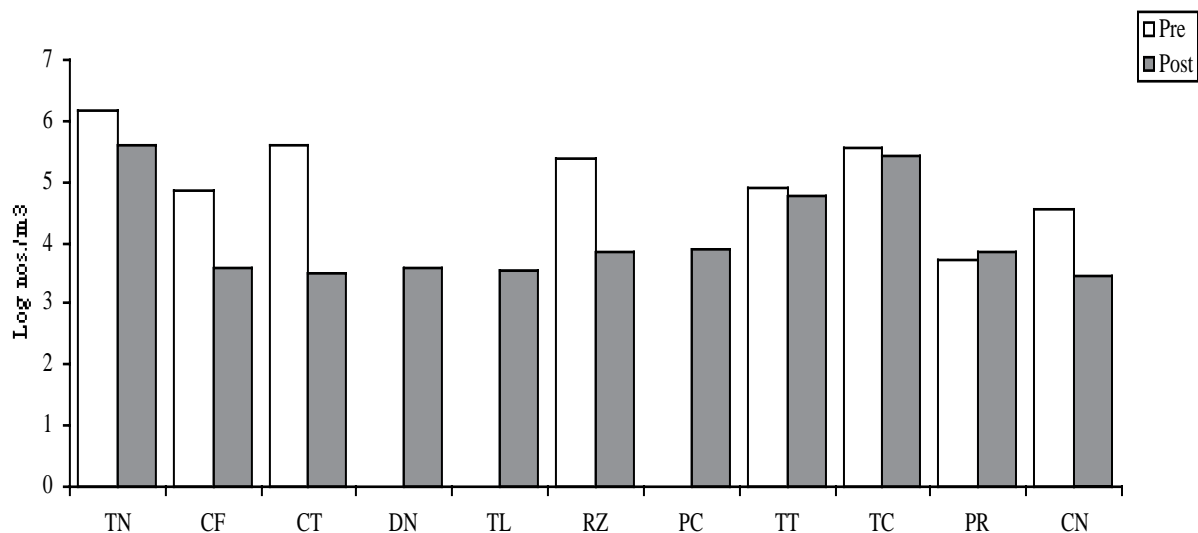


Fig. 8 Cell densities (log nos./m³) of various microplankton species at stations in Eastern Sabah waters of the South China Sea during pre and post monsoon seasons. (TN-total cell, CF-*Ceratium furca*., CT-*Chaetoceros macroceros*, DN-*Dinophysis*, TL-*Thaliassionema nitzschoides*, RZ-*Rhizosolenia*, TT-*T. thiebautii*., TC-*Trichodesmium erythraeum*, PR-*Protoperdinium* sp., CN-Copepod nauplii).

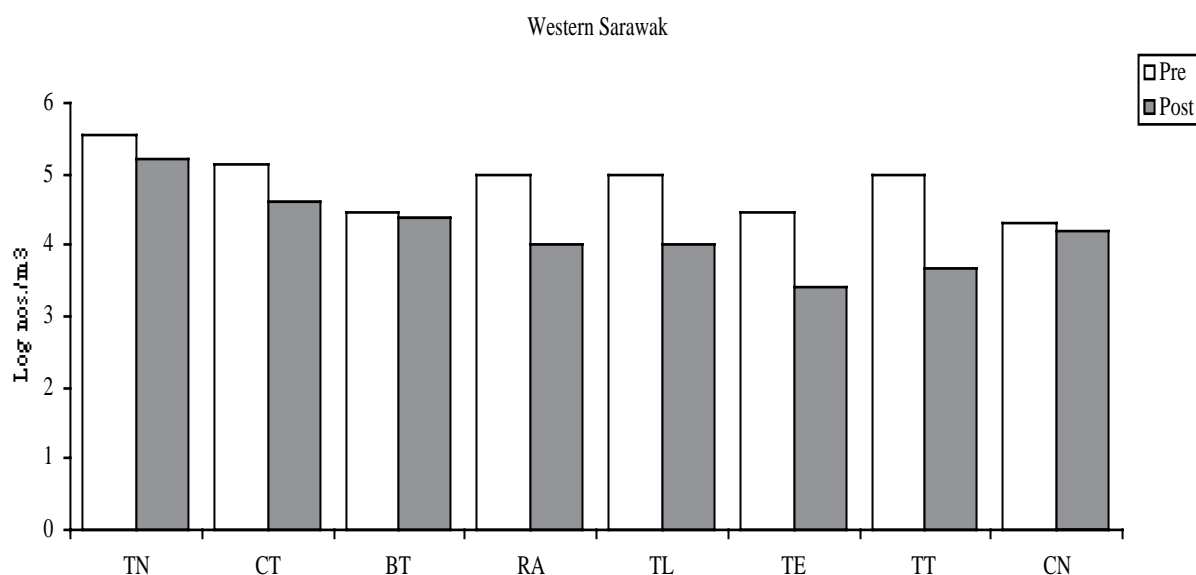


Fig. 9 Population densities (log nos./m³) of various microplankton species at stations in Western Sarawak Waters of the South China Sea during pre and post monsoon seasons. (TN-total cell, CT-*Chaetoceros lorenzianum*, BT-*Bacteriastrum delicatulum*, RA-*Rhizosolenia alata*, TL-*Thalassionema frauenfeldii*, TE-*Trichodesmium erythraeum*, TT-*T. thiebautii* sp., CN-Copepod nauplii).

group C (*Richelia* sp., *Rhizosolenia clevei*, *Peridinium* sp., *Pleurosigma* sp., *Climacodium* sp.); group D (*Trichodesmium erythraeum*, *T. thiebautii*, *Protoperdinium* sp., *Hemialus* sp.); group E (*Rhizosolenia habatata*, *Globigerina* sp., *Chaetoceros decipiens*); group F (*Chaetoceros comosum*, *Ceratium fusus*, *C. teres*).

Earlier studies by Shamsudin *et al.* 1987 in the Malaysian waters of the South China Sea showed that the microphytoplankton from 16 oceanographic stations consisted predominantly of diatoms and blue green algae. The bulk of the diatom species consisted of *Chaetoceros*, *Rhizosolenia*, *Melosira*, *Thalassiothrix*, *Dactyliosolen* and *Guinardia*. Another diatom species, *Planktoniella* was present only at stations further offshore from the coast. However, other diatom species which were also present included those species of *Bacteriastrum*, *Asterionella*, *Fragilaria*, *Nitzschia*, *Skeletonema*, *Coscinodiscus* and *Pleurosigma*. More than 30 major species of diatom have been identified. The genera *Coscinodiscus*, *Chaetoceros* and *Rhizosolenia* were found to contain a wide range of species. The Cyanophyta comprised of only a few species among which *Trichodesmium thiebautii* and *T. erythraeum* were present in abundant.

Other studies of microplankton in Malaysian waters including the Straits of Malacca had been conducted by Sewell (1933), Winstead (1961), Pathansali (1968), Chua & Chong (1973), Shamsudin (1987, 1993, 1994, 1997) and Shamsudin & Shazali (1991). Most of these studies were carried out at certain predetermined time and location; however, the present study was carried out during the pre and postmonsoon periods. An increase in the diversity value could be due to an increased number of species or even distribution of individuals per species as described by Gray (1981). In reality, such community organisation is constantly acted on by biological and physical factors in many different ways to produce, perhaps a different organisation in the future as a response to such environmental changes. When a bloom occurs, only a few microplankton species will predominate and thus effect or influence the number of species or the even distribution of individual species.

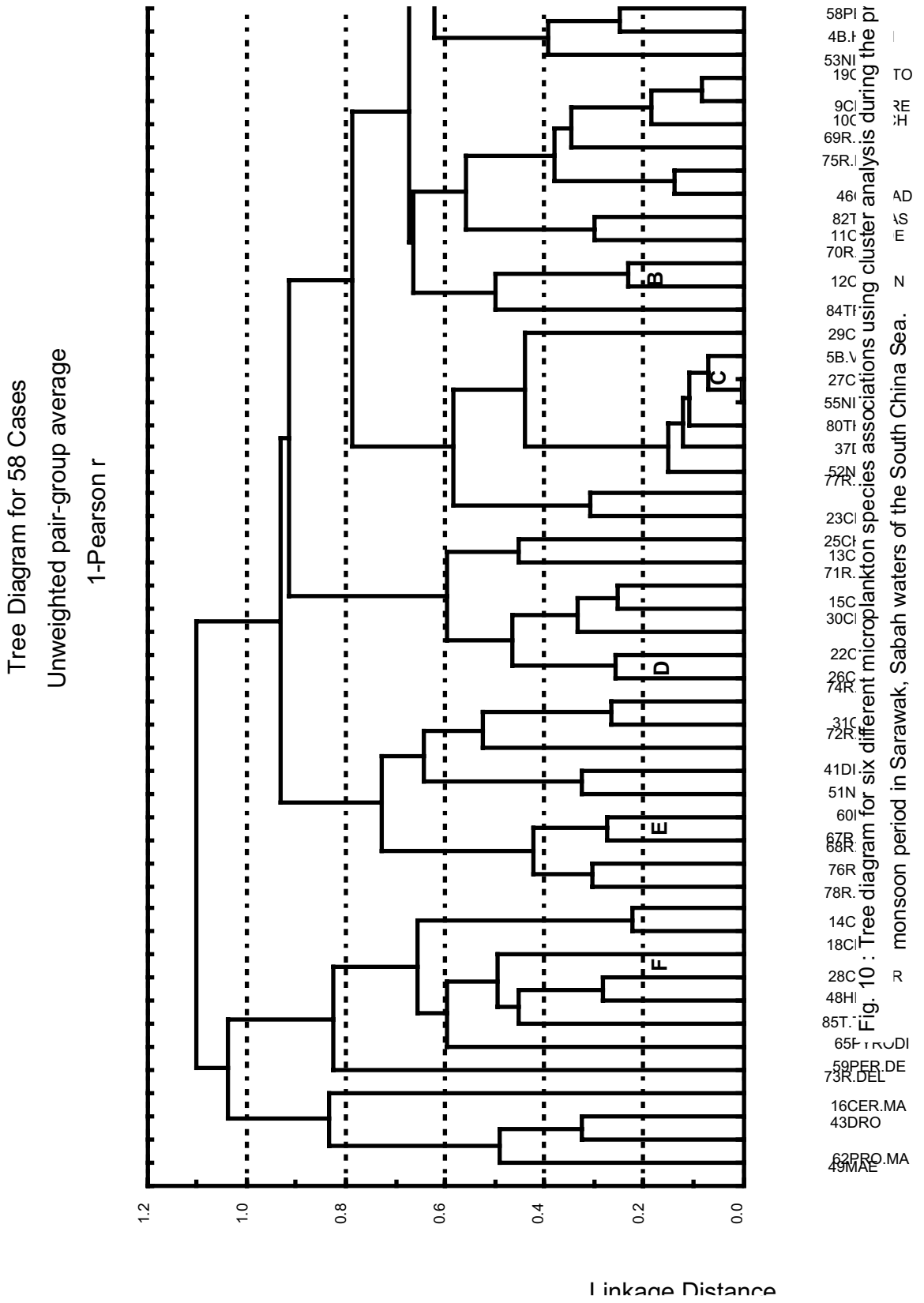


Fig. 10

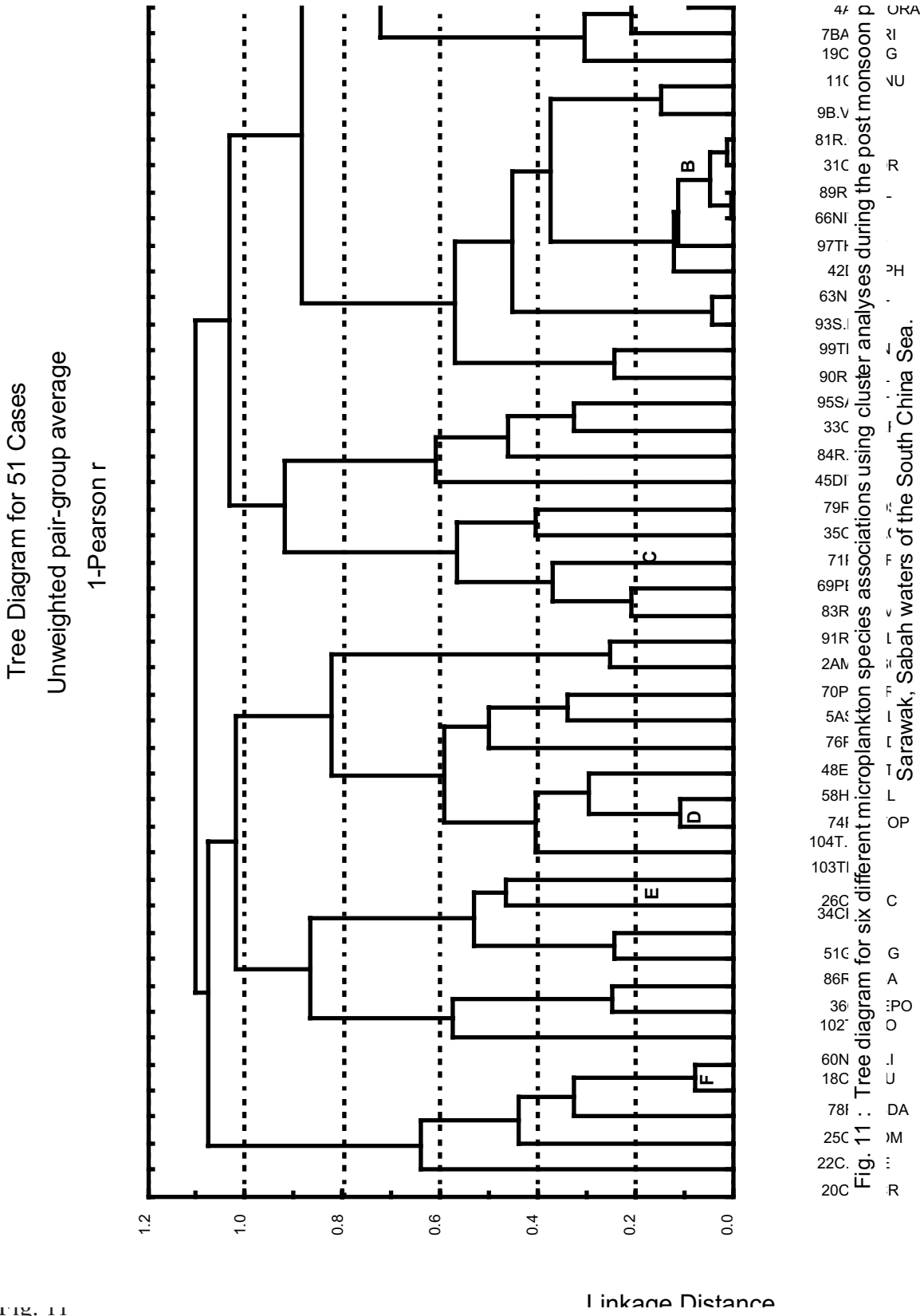


Fig. 11

Microzooplankton population

Microzooplankton species consisted of more than 20 different groups with several dominant species namely, copepod nauplii (> 50% of the total microzooplankton count); Chaetognatha larvae (5%), Ciliophora (4%) and Foraminifera (2-3%) (see Appendix 2). The Ciliophora consisted of a few genera (*Tintinnopsis*, *Distephanus*, *Tintinnus*, *Favella*, *Dictyocha*, *Tomopteris*, *Xystonella*, *Xystonellopsis*, *Codonellopsis*) while Foraminifera consisted of *Globigerina* and *Tretomphalus* species (considered to be indicator tropical species). *Amphisolenia* (Peridinidae) and *Ceratocorys* species were detected in considerable amount in nearshore stations. Numerous *dinoflagellates* (*Dinophysis favus*, *D. norvegicus*, *D. triposolnia*, *Ornithocercus* sp., *Peridinium brochii*, *P. depressum*, *P. subpyriformes*, *Podolampas* sp., *Prorocentrum* sp., *Pyrocystis fursiformis*, *P. lunula*, *Pyrophacus horologium*, *Rhabdonella* sp.) were found in middle Sarawak waters of the South China Sea.

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Table 1 The number of species in the genera of the microplankton population in Sabah and Sarawak waters of the South China Sea during the study period (* Dominant)

Genus	Number of species	Genus	Number of species
1 Bacillariophyceae (Diatom)		2 Dinophyceae	
<i>Asteromphalus</i>	2	<i>Goniodoma</i>	2
* Bacillaria	1	* <i>Gonyaulax</i>	4
<i>Compylodyscus</i>	4	* <i>Noctiluca</i>	1
<i>Cerataulina</i>	3	* Ornithocercus	5
<i>Climacodium</i>	2	<i>Palacroma</i>	5
<i>Corethron</i>	1	* Podolampas	4
<i>Dactyliosolen</i>	2	<i>Prorocentrum</i>	3
<i>Ditylum</i>	2	* Protoperidinium	5
* <i>Eucampia</i>	2	<i>Pyrophalus</i>	2
* <i>Fragilaria</i>	1		
* <i>Guinardia</i>	3	3 Cyanophyceae	
* <i>Gyrosigma</i>	4	Trichodesmium	2
* <i>Hemiaulus</i>	3		
* <i>Hemidiscus</i>	2	4 Dictyochaceae	
* <i>Lauderia</i>	2	<i>Dictyocha</i>	4
<i>Leptocylindrus</i>	2	<i>Procentrum</i>	1
* <i>Nitzschia</i>	8	Pyrocystis	3
<i>Odentella</i>	2		
* <i>Planktoniella</i>	2	5 Microzooplankton	
<i>Pseudoguinardia</i>	2	<i>Globigerina</i>	1
* Skeletonema	1	<i>Codonella</i>	1
* <i>Thalassiosira</i>	5	Tintinnopsis	2
<i>Triceratium</i>	1	Favella	1
		<i>Tintinnus</i>	1
2 Dinophyceae			
<i>Alexandrium</i>	2	6 Larvae/nauplii	
<i>Amphisolenia</i>	4	<i>Chaetognatha</i>	-
* Ceratium	28	<i>Ostracoda</i>	-
* Ceratocorys	2	<i>Siphonophora</i>	-
* <i>Dinophysis</i>	5	<i>Gastropod</i>	-

Table 2. Dominant microplankton species at various sectors in Sabah and Sarawak waters of the South China Sea during the study period.

Sector	Monsoon	
	Pre	Post
Sarawak coastal waters (SCW)	<i>Trichodesmium erythraeum</i> <i>Ceratium furca</i> <i>Rhizosolenia stolterfothii</i> <i>Chaetoceros lorenzianus</i>	<i>Trichodesmium erythraeum</i> <i>Thalassionema fraunfeldii</i> <i>Thalassionema nitzschooides</i> <i>Rhizosolenia stolterfothii</i> <i>Bacteriastrium varians</i> <i>Chaetoceros lorenzianum</i>
Western Sarawak waters (WSW)	<i>Trichodesmium erythraeum</i> <i>Rhizosolenia alata</i> <i>Coscinodiscus</i> sp.	<i>Trichodesmium thiebautii</i> <i>Trichodesmium erythraeum</i> <i>Thalassionema nitzschooides</i> <i>Thalassionema delicatulum</i> <i>Rhizosolenia alata</i>
Offshore Sarawak waters (ESW)	<i>Trichodesmium erythraeum</i> <i>Rhizosolenia alata</i> <i>Coscinodiscus</i> sp. <i>Peridinium</i> sp.	<i>Trichodesmium erythraeum</i> <i>Trichodesmium thiebautii</i> <i>Rhizosolenia hebatata</i> <i>Bacteriastrium hyalinum</i> <i>Chaetoceros didymum</i>
Eastern Sabah waters (ESW)	<i>Trichodesmium erythraeum</i> <i>Rhizosolenia alata</i>	<i>Trichodesmium erythraeum</i> <i>Ceratium furca</i> <i>Peridinium depressum</i> <i>Protocentrum</i> sp. <i>Rhizosolenia alata</i>
Intermediate Middle Sarawak waters (IMSW)	<i>Trichodesmium erythraeum</i> <i>Rhizosolenia alata</i> <i>Pleurosigma</i> sp. <i>Chaetoceros lorenzianus</i>	<i>Trichodesmium erythraeum</i> <i>Trichodesmium thiebautii</i> <i>Rhizosolenia hebatata</i> <i>Bacteriastrium hyalinum</i> <i>Bacteriastrium varians</i>

Table 3. Species association in Sabah and Sarawak waters of the South China Sea during the pre and post monsoon period

Group	Monsoon	
	Pre	Post
A	<i>Chaetoceros coarctatum</i> <i>Thalassionema fraeunfeldii</i> <i>Bacteriastrum delicatulum</i>	<i>Ceratocorys</i> sp. <i>Bacteriastrum hyalinum</i> <i>Nitzschia frigida</i>
B	<i>Trichodesmium erythraeum</i> <i>Ceratium arauatum</i>	<i>Thalassionema fraeunfeldii</i> <i>Nitzschia seriata</i> <i>Rhizosolenia stolterforthii</i> <i>Chaetoceros lorenzianum</i>
C	<i>Thalassionema fraeunfeldii</i> <i>Chaetoceros lorenzianum</i>	<i>Richelia</i> sp. <i>Rhizosolenia clevei</i> <i>Peridinium</i> sp. <i>Pleurosigma</i> sp. <i>Climacodium</i> sp.
D	<i>Rhizosolenia hebatata</i> <i>Chaetoceros lacinosus</i> <i>Chaetoceros decipiens</i> <i>Ceratium longissinum</i>	<i>Trichodesmium erythraeum</i> <i>Trichodesmium thiebautii</i> <i>Protoperidinium</i> <i>Hemialus</i> sp.
E	<i>Rhizosolenia styliformis</i> <i>Rhizosolenia robusta</i> <i>Rhizosolenia bergonii</i> <i>Rhizosolenia alata</i>	<i>Rhizosolenia hebatata</i> <i>Globigerina</i> sp. <i>Chaetoceros decipiens</i>
F	<i>Trichodesmium thiebautii</i> <i>Chaetoceros peruvianum</i> <i>Ceratium teres</i> <i>Ceratium fusus</i>	<i>Chaetoceros comosum</i> <i>Ceratium fusus</i> <i>Ceratium teres</i>

Appendix 1 The taxonomic list of Microzooplankton identified from Sarawak and Sabah waters South China Sea (* Dominant)

<p>1 Class, Cyanophyceae; Order Hormogoneae; Family Oscillatoriaceae; * <i>Trichodesmium erythraeum</i> Ehrenberg <i>T. thiebautii</i> Gom.</p>	<p><i>C. setaceum</i> Jorg <i>C. siamense</i> Ostenfeld <i>C. sumatranum</i> Karsten <i>C. tetrastichon</i> Cleve <i>C. tripos</i> Nitsch <i>C. weissflogii</i> Schutt</p>
<p>2 Pylum Bacillariophyceae (Diatom) <i>Actinophychus undulatus</i> Ralfs <i>Actinocyclus</i> Ehrenberg <i>Asterolampra marylandica</i> Ehrenberg <i>Asteromphalus elegans</i> Greville <i>A. heptactis</i> Ralfs <i>A. flabellatus</i> Greville <i>Bacillaria paxillifera</i> O.F. Muller <i>Bacteriastrium comossum</i> Pavillard * <i>B. delicatulum</i> Cleve <i>B. elegans</i> Pavillard <i>B. elongatum</i> Cleve * <i>B. hyalinum</i> Lauder <i>B. mediaterraneum</i> Pavillard <i>B. minus</i> Lauder * <i>B. varians</i> Lauder <i>Biddulphia dubia</i> Cleve <i>B. longicrucia</i> Greville * <i>B. mobilensis</i> Bailey <i>B. regia</i> Ostenfeld <i>B. sinensis</i> Grevillae <i>Campylodiscus biangulatus</i> Hantsch <i>C. daemelianus</i> Grun <i>C. echeneis</i> Ehrenberg <i>C. ornatus</i> Grun <i>C. undulatus</i> Grevillae <i>Cerataulina Bergonii</i> <i>C. Compacta</i> Ostenfeld <i>C. pelagica</i> (Cleve) Hendey <i>C. coarctatum</i> Lauder <i>Chaetoceros affinis</i> Lauder <i>C. brevis</i> Schutt <i>C. compressum</i> Lauder <i>C. constrictum</i> Gran <i>C. costatus</i> Pavillard * <i>C. curvisetum</i> Cleve <i>C. dadayi</i> Pavillard <i>C. debile</i> Cleve * <i>C. decipiens</i> Cleve <i>C. densum</i> Cleve <i>C. denticulatum</i> Lauder <i>C. decipiens</i> Cleve * <i>C. didymum</i> Ehrenberg * <i>C. distans</i> Ehrenberg * <i>C. diversus</i> Cleve <i>C. hispidum</i> Brightwell <i>C. indicum</i> Koosten * <i>C. lacinosus</i> Schutt <i>C. latderi</i> Rafts <i>C. lauderi</i> Reefs <i>C. leavis</i> Leuduger - Fortimorel <i>C. messanensis</i> Castracane <i>C. paradoxum</i> Cleve <i>C. pendulus</i> Karsten * <i>C. peruvianum</i> Brightwell * <i>C. pseudocurvisetum</i> Mangin</p>	<p><i>Climacodium biconcavum</i> Cleve <i>C. frauenfeldianum</i> Grunow <i>Corethron hystrix</i> Henden <i>C. pelagicum</i> Brun <i>Coscinodiscus asteromphalus</i> Ehrenberg * <i>C. concinus</i> W. Smith * <i>C. centralis</i> Grunow * <i>C. curvatulus</i> Grunow * <i>C. debilis</i> Ehrenberg * <i>C. gigas</i> Ehrenberg <i>C. granii</i> Gough <i>C. janischii</i> Schmidt * <i>C. jonesianus</i> (Greville) Ostenfeld <i>C. lineatus</i> Ehrenberg <i>C. marginatus</i> Ehrenberg <i>C. nitidus</i> Gregory <i>C. nobilis</i> Grunow <i>C. nodulifer</i> Schmidt <i>C. oculus rividis</i> Ehrenberg <i>C. perforatus</i> Ehrenberg <i>C. radiatus</i> Ehrenberg <i>C. Rothii</i> Grunow <i>C. stellaris</i> Roper <i>C. subtilis</i> Ehrenberg <i>C. weilesii</i> Gran & Angst <i>Cylindrotheca closterium</i> Ehrenberg <i>Dactyliosolen blavyanus</i> H. Peragallo <i>D. fragilissimum</i> (Bergon) Hasle <i>Detonula pumila</i> (Castracane) Gran <i>Ditylum brightwellii</i> (West) Grunow <i>D. sol</i> Grunow <i>Eucampia cornuta</i> (Cleve) Grunow <i>E. zodiacus</i> Ehrenberg <i>Fragilaria intermedia</i> Grunow <i>Guinardia cylindrus</i> (Cleve) Hasle <i>G. flaccida</i> (Castracane) H. Peragallo <i>G. striata</i> Stolteriotn Hasle <i>Gossleriella tropica</i> Schutt <i>Gyrosigma acuminatum</i> Rabh <i>G. balticum</i> Cleur <i>G. Strigile</i> Smith <i>Halicotheca thamensis</i> Grunow <i>Hemiaulus hauckii</i> Grunow <i>H. indicus</i> Karsten <i>H. membranacea</i> Cleve <i>H. sinensis</i> Greville * <i>Hemidiscus cuneiformis</i> Wallich (Indicator sp.) <i>H. hardmanianus</i> <i>Lauderia annulata</i> Gran <i>L. borealis</i> Gran <i>Leptocylindrus danicus</i> Cleve <i>L. mediterraneus</i> (H. Peragallo) Hasle <i>Lithodesmium undulatum</i> Ehrenberg <i>Navicula</i> sp. * <i>Nitzschia closterium</i> W. Smith</p>

Appendix 1 Continue

<i>N. closterium</i> W. Smith	<i>Amphidoma steini</i> Schill
<i>N. hungarica</i> Grun	<i>Amphisolenia bidentata</i> Schroder
<i>N. lanceolata</i> W. Smith	<i>A. thrinax</i> Schutt
<i>N. longissima</i> Gran	<i>A. globifera</i> Stein
<i>N. longissima</i> var. <i>reversa</i> W. Smith	<i>A. schnauinsianaii</i> Lemmermann
<i>N. paradoxa</i> Gmelin	Ceratium axiale Kofoid
<i>N. pacifica</i> Cupp	<i>C. arietinum</i> Cleve
<i>N. plana</i> W. Smith	* <i>C. breve</i> Schroder
<i>N. pungens</i> Cleve	<i>C. biceps</i> Gourret
<i>N. seriata</i> Cleve	<i>C. belone</i> Cleve
<i>N. sigma</i> W. Smith	<i>C. condillans</i> Jorgensen
<i>N. sigma</i> var. <i>intercedens</i> Grun	<i>C. candelabrium</i> Ehrenberg Stein
<i>N. spectavilis</i> Ralfs	<i>C. contortum</i> Gourret
<i>N. vitrea</i> Norman	<i>C. carriense</i> Gourret
<i>N. bicapitata</i> Cleve	<i>C. declinatum</i> (Karsten) Jorgensen
Odontella mobiliensis (Bailey) Grunow	* <i>C. deflexum</i> (Kofoid) Jorgensen
<i>O. sinensis</i> (Greville) Grunow	<i>C. dens</i> Ostenfeld & Schmidt
Planktoniella blanda A. Schmidt	<i>C. falcatum</i> (Kofoid) Jorgensen
<i>P. sol</i> (Wallich) Schutt	<i>C. furca</i> Ehrenberg
Pleurosigma affine Gran	<i>C. fusus</i> Ehrenberg
<i>P. angulatum</i> W. Smith	* <i>C. gibberum</i> Gourret
<i>P. coompactum</i> Grew	* <i>C. gravidum</i> Gourret
* <i>P. elongatum</i> W. Smith	<i>C. hexacanthum</i> Gourret
<i>P. fasciola</i> W. Smith	<i>C. horridum</i> (Cleve) Gran
<i>P. intermedium</i> W. Smith	<i>C. inflatum</i> (Kofoid) Jorgensen
<i>P. nicobaricum</i> Gran	<i>C. kofoidii</i> Jorgensen
<i>P. Normanii</i> Ralfs	<i>C. longissimum</i> Gran
<i>P. pelagicum</i> Perag	<i>C. limulus</i> Gourret
<i>P. rectum</i> Donkim	<i>C. lunula</i> (Schimpe) Jorgensen
<i>P. rigidum</i> Brun	* <i>C. macroceros</i> (Ehrenberg) Vanhoff
<i>P. salinarum</i> Gran	* <i>C. massiliense</i> (Gourret) Karsten
Pseudoguinaridia recta Von Stesen	* <i>C. platycorne</i> Daday
<i>P. pungens</i> Grunow & Cleve Hasle	* <i>C. pentagonum</i> Gourret
* Rhizosolenia acuminata Gran	<i>C. pulchellum</i> Schroder
* <i>R. alata</i> Brightwell	<i>C. symmetricum</i> Paviilard
<i>R. bergonii</i> H. Peragallo	* <i>C. teres</i> Kofoid
<i>R. clevei</i> Ostenfeld	<i>C. trichoceros</i> (Ehrenberg) Kofoid
<i>R. castracanei</i> H. Perag	<i>C. tripos</i> (O.F. Muller) Nitzsen
<i>R. curvata</i> Zacharias	<i>C. vulture</i> Cleve
* <i>R. calcar-avis</i> M. Schutze	Ceratocorys norrida Stein
<i>R. formosa</i> H. Peragallo	<i>C. horrida</i> Stein
<i>R. cylindrus</i> Cleve	<i>C. gourreti</i> Paulsen
<i>R. hyaline</i> Ostenfeld	Corythodinium resseratum Stein
<i>R. delicatula</i> Cleve	Loeblich Jr. & Loebien
<i>R. imbricata</i> Brightwell	Dinophysis homunculus Stein
<i>R. hesetata</i> Gran	<i>D. caudata</i> Sabille - Kent
<i>R. robusta</i> Norman	<i>D. hastata</i> Stein
<i>R. delicatula</i> Cleve	<i>D. infundibula</i> Schiller
<i>R. setigara</i> Brightwell	<i>D. miles</i> Cleve
<i>R. styliformis</i> Brightwell	<i>D. ovum</i> Schutt
* Skeletonema costatum (Greville) Cleve	<i>D. schuettii</i> Murray & Whitting
Stephanopyxis palmeriana Greville	<i>D. tripos</i> Gourret
Striatella sp.	Diplopsalis lenticulata Berg
Suriella sp.	Goniodoma polyedricum Pouchet
* Thalassionema frauenfeldii Grunow	<i>G. spaericum</i> Murr. & Whitt
<i>T. nitzschioides</i> Grunow	Gonyaulax digitale (Pouchet) Kofoid
Thalassiosira bingensis Takano	<i>G. gluptorhynchus</i> Murray & Whitting
<i>T. dipporocyclus</i> Hasle	<i>G. polygramma</i> Stein
<i>T. eccentrica</i> (Ehrenberg) Hasle	<i>G. spinifera</i> Clapareda & Lachmann
<i>T. oestrupii</i> (Ostenfeld) Hasle	Gynmodinium sp.
* <i>T. subtilis</i> (Ostenfeld) Gran	Gyrodinium sp.
Triceratium favus Ehrenberg	Kofoidinium sp.
3 Phylum Dinophyceae	Noctiluca scintillans Macartney
(Dinoflagellate)	Ornithocercus magnificus Stein
Family : Peridiniidae	<i>O. thumii</i> A. Schmidt
Alexandrium fraterculus (Balech)	<i>Pxytoxum scolopax</i> Stein
<i>A. tamiyavanichi</i> Balech	<i>O. milneri</i> Gran

Appendix 1 Continue

<i>O. tessellatum</i> Stein	<i>P. murrayi</i> (Kofoid) Balech
<i>Phalacroma acutoides</i> Balech	* <i>P. oceanicum</i> (Vanhoff) Balech
<i>P. doryphorum</i> Stein	<i>P. okamurai</i> (Abe') Balech
<i>P. favus</i> Kofoid & Micherner	<i>P. ovum</i> (Schiller) Balech
<i>P. mitra</i> Schutt	<i>P. pallidum</i> (Ostenfeld) Balech
<i>P. parvulum</i> Schutt	<i>P. paulseni</i> (Pavillard) Balech
<i>P. rapa</i> Stein	<i>P. Pellucidum</i> Bergn
<i>P. rudgei</i> Murray & Whitting	<i>P. puanerense</i> (Schreaser) Balech
<i>Podolampas bipes</i> Stein	<i>P. spinuosum</i> (Schiller) Balech
<i>P. elegans</i> Schutt	<i>P. stenii</i> (Jorgensen)
<i>P. palmipes</i> Stein	<i>P. thorianum</i> (Paulsen) Balech
<i>P. spinifera</i> Okamura	<i>Pyrophacus horologium</i> Stein
<i>Prorocentrum compressum</i> (Bailey)	<i>P. stein</i> (J. Schiller) Wall & Dale
<i>P. micans</i> Ehrenberg	<i>Scripsiella trochoidea</i> (Stein) Balech
<i>P. sigmoides</i> Bohm	4 Family : Dictyochaceae
* <i>Protoceratium spinulosum</i>	(Phylum Protozoa)
<i>Protoperidinium conicum</i> (Gran)	Class : Mastogophora
* <i>P. brochii</i> Balech	Order : Chrysomonadina
<i>P. crassipes</i> (Kofoid) Balech	<i>Dictyocha fibula</i> Ehrenberg
<i>P. depressum</i> (Bsiley) Balech	<i>D. fibula</i> var stapedia Heack
<i>P. diabolus</i> (Cleve) Balech	<i>D. fibula</i> var major Rampi
<i>P. divergens</i> (Ehrenberg) Balech	Family : Procentridae
<i>P. elegans</i> (Cleve) Balech	<i>Procentrum micans</i> Ehrenberg
<i>P. globulum</i> (Stein) Balech	Family : Phytodinidae
* <i>P. grande</i> (Kofoid) Balech	* <i>Pyrocystis elegans</i> Murray
<i>P. hirobis</i> (Abe') Balech	(Indicator sp.)
<i>P. latispinum</i> (Mangin) Balech	<i>P. fusiformis</i> Murray
<i>P. leonis</i> (Pavillard) Balech	<i>P. hamulus</i> var imacqualis Schrober
	<i>P. noctulica</i> Murray